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## Conformationally restricted macrocyclic analogues of combretastatins

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**Abstract**—New analogues of combretastatins have been evaluated as inhibitors of tubulin polymerization. These compounds present a macrocyclic structure, in which the *para* positions of the aromatic moieties have been linked by a 5- or 6-atoms chain, in order to produce a conformational restriction. This could contribute to determine the active conformation for these ligands. Such a conformational restriction and/or the steric hindrance makes them less potent inhibitors than the model compound CA-4. © 2007 Elsevier Ltd. All rights reserved.

Microtubules are structures formed by polymerization of  $\alpha\beta$ -tubulin dimers. They are essential to many cellular processes, such as maintenance of cellular shape, intracellular transport, or mitotic spindle assembly during cell division. Inhibition of microtubule formation leads to mitotic arrest and promotes vascular disruption in angiogenic vessels through activation of the RhoA and/or the VE-cadherin/ $\beta$ -catenin pathways. Tubulinbinding agents such as colchicine or vinblastine display both effects, but in these two cases the vascular shutdown occurs at concentrations close to the maximum tolerated dose.  $^3$ 

Combretastatin A-4 (CA-4, Fig. 1) is one of the most potent inhibitors of tubulin polymerization, binding at the colchicine site on the protein. It is currently in phase I/II clinical trials as its phosphate prodrug<sup>4</sup> due to its antimitotic and antiangiogenic properties. Other members of the combretastatin family also inhibit tubulin polymerization, such as deoxycombretastatin,<sup>5</sup> combretastatin,<sup>6</sup> and related dihydroxy and dioxolane derivatives (Fig. 1),<sup>7</sup> among others. In the two latter, the configuration of the stereogenic carbons has proved to be essential for antitubulin activity.

tural scaffolds and spatial dispositions of the aromatic rings.

Many combretastatin analogues have been prepared in which the phenyl rings are locked in the *cis* orientation by the formation of small cycles on the bridge, in an attempt to prevent them from isomerizing.<sup>8,10</sup>

As part of our research directed at the synthesis and evaluation of new antimitotic agents, we have designed

The structural requirements for a potent antitubulin activity are those present in CA-4.8 A 3,4,5-trimethoxy-

phenyl and a 4-methoxy-3-X-phenyl (being X = H, OH,

NH<sub>2</sub> and their aminoacyl, phosphate, or other solubiliz-

ing derivatives) separated by a 2-atoms bridge with cis

disposition are the structural features for the most po-

tent compounds. The aromatic rings show a non-copla-

nar disposition. However, the conformation of CA-4

bound to tubulin is not known. Colchicine and podo-

phyllotoxin show different spatial arrangement of the

aromatic rings when bound to tubulin<sup>9</sup> and, despite this,

both of them are potent inhibitors of polymerization (IC<sub>50</sub> = 0.8–3.3 and 0.3–3  $\mu$ M, respectively). This fact has been attributed to the flexibility of the binding site,

which can accommodate ligands bearing different struc-

new analogues of combretastatins in which a macrocycle is formed by linking the *para* positions of both aromatic rings, through the corresponding oxygen or nitrogen atoms. This restriction generates new *cis*-locked analogues irrespective of the bridge attracture. Conformed

logues, irrespective of the bridge structure. Conformational restriction by macrocycle formation on bioactive

*Keywords*: Combretastatins; Conformational restriction; Macrocycles; Tubulin polymerization inhibition.

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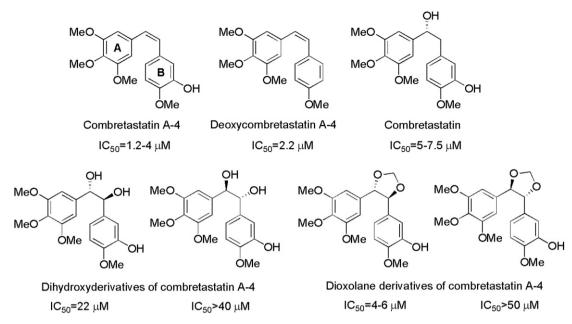


Figure 1. Structure and potency as inhibitors of tubulin polymerization of combretastatin A-4, deoxycombretastatin A-4, combretastatin and related diols and dioxolanes.

compounds has been used to increase binding affinity, selectivity, and metabolic stability. Recently, the effect of macrocyclization and stereochemistry on the biological activity of small molecules has been quantitatively measured.<sup>11</sup>

Two linkers with different length (5 or 6 atoms) were chosen to afford a different degree of conformational restriction by varying the macrocycle size. Thus, 3-oxapentamethylene or hexamethylene linkers were employed, giving rise to Families I and II. This modification involves a minor increment of molecular size (CH<sub>2</sub>–O–CH<sub>2</sub> or (CH<sub>2</sub>)<sub>4</sub>, see Fig. 2), restricted to the space between the aromatic rings and thus without affecting the more external surface.

The substitution pattern of both aromatic rings was selected so to mimic the structural features that confer high potency to combretastatins. Thus, 3,5-dimethoxy groups were attached to phenyl ring A (the 4-methoxy group being part of the linker). For ring B, three possibilities were envisaged. First, keep it without substitution, as in deoxycombretastatin A-4. Second, attach a

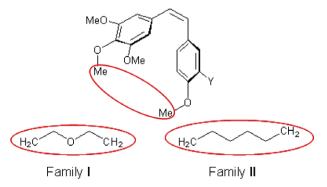


Figure 2. Structure of the linkers.

hydroxyl group at position 3, as in combretastatin A-4 (the 4-methoxy group also being part of the linker in both cases). And third, replace the phenyl ring by a more rigid 5-indolyl system, which had produced potent compounds as surrogate for ring B.<sup>12</sup>

As the bridge between the rings, the analogues bear the double bond present in combretastatin A-4, the corresponding glycol (*cis* or *trans*) or its derivatives (acetate or dioxolane). We expected that the conformational restriction imposed on these compounds by the formation of the macrocycle (also influenced by the substitution pattern on the rings<sup>13</sup> and on the bridge<sup>14</sup>) could make them to adopt optimal binding geometries, so they could show an increase in tubulin binding potency. The results will then shed light on which is the active conformation of combretastatins when bound to tubulin.

The synthesis of this type of compounds has been reported previously by us. <sup>13,14</sup> Briefly, olefins and glycols were prepared by means of an intramolecular McMurry reaction of the dialdehydes. These are in turn synthesized by double Mitsunobu reaction starting from the corresponding aromatic aldehydes and the appropriate linker diol (diethyleneglycol for Family I or 1,6-hexanediol for Family II) (Fig. 3). The synthesized compounds are shown in Figure 4.

These macrocyclic combretastatin analogues were evaluated as tubulin polymerization inhibitors. The assay conditions are as previously reported.  $^{12}$  The compounds were usually assayed at 20 or 40  $\mu M$  (some analogues were assayed at lower concentration due to scarce aqueous solubility) and the results expressed as percentage of polymerization of the control (Table 1). It should be noted that, in order to compare the potencies of the analogues, some values of ITP must be extrapolated due to the different concentrations assayed.

Figure 3. Retrosynthetic analysis for the designed compounds.

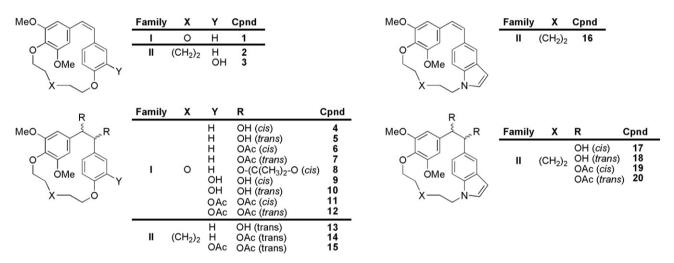


Figure 4. Compounds evaluated as inhibitors of tubulin polymerization.

The assayed analogues show a moderate to low potency related to the reference compound CA-4. The best inhibitors of tubulin polymerization belong to the group of the olefin-bridged analogues (2 and 16), as seen in non-cyclized model compounds. The conformational analysis for these analogues reveals a severe conformational restriction compared to the parent combretastatins. 13,14 They show a disposition between the aromatic rings less orthogonal than that showed by tubulin-bound colchicine and podophyllotoxin. The rotation of the rings is greatly hampered by the presence of the linker and the substituents on the rings. Such a restriction is somewhat lower for compounds of Family II, in which a longer 6-atoms linker allows more flexibility and thus a larger conformational space for the aromatic rings. This would account for the highest potency of compound 2 (ITP = 60% at  $40 \mu M$ ). When the linker is shorter (3-oxapentamethylene), none of the analogues showed any relevant activity (compare 1 with the most potent 2).

The presence of a  $\sigma$ -bond bridge in the glycols and derivatives modifies the dihedral angles between the ring

axes related to the olefins. They can adopt two defined conformations, exchangeable by rotation of such single bond. The absence of significant antitubulin activity for these analogues would imply that neither of those conformations are able to fit into the binding site of tubulin, regardless of the substitution pattern on the rings, the configuration of the stereogenic carbons or the length of the linker.

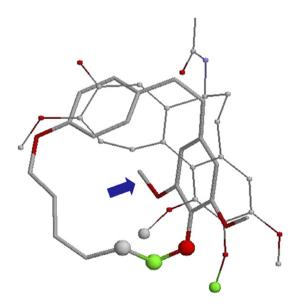
Therefore, it seems that the conformational restriction imposed to these combretastatin analogues by the formation of a macrocycle prevents them from adopting a suitable conformation for a productive binding that leads to potent antitubulin activity. It cannot be ruled out that the low potency of these compounds could be also accounted for by the steric hindrance caused by the presence of the linker between the rings. Because combretastatin analogues with bulky substituents on ring B (hydroxyethoxy, pyrrolylethoxy, imidazolylethoxy) keep high to moderate potency, 15 the linker may be detrimental if the inter-ring space is required by the protein to adapt itself to the combretastatin geometry.

**Table 1.** Inhibition of tubulin polymerization (ITP) for compounds 1–

Compound	Assayed concentration (μM)	Inhibition of tubulin polymerization (%)
1	30	0
2	40	60
3	40	0
4	40	0
5	40	0
6	40	0
7	40	0
8	40	0
9	40	11
10	40	1
11	40	0
12	40	0
13	40	4
14	40	0
15	40	2
16	10	19
17	20	11
18	20	9
19	20	9
20	20	8
CA-4	3	50

ITP values represent means of three experiments.

On the other hand, the superposition of the structures of tubulin-bound DAMA-colchicine<sup>9</sup> and compound 2<sup>16</sup> shows that part of the linker could fill the space occupied by two of the methoxy groups of colchicine (Fig. 5). Then, the second methoxy group on ring A of the macrocyclic analogues would be unnecessary or even detrimental for activity. Moreover, the conformational freedom of ring A would also be somewhat higher due to a lower steric hindrance. Therefore, a new family of analogues with only one methoxy group on ring A is



**Figure 5.** Superposition of tubulin-bound DAMA-colchicine (thin sticks) and compound **2** (bold sticks). The balls represent methoxy groups of colchicine that could be replaced by part of the linker. The blue arrow represents the unnecessary methoxy group on compound **2**.

being prepared and the results of their biological evaluation will be reported in due course.

In summary, we have evaluated a number of new analogues of combretastatins as inhibitors of tubulin polymerization. These compounds have a macrocyclic structure generated by the presence of a 5- or 6-atoms linker between the *para* heteroatoms of the aromatic systems in order to impose a conformational restriction. Such a restriction prevents them from adopting a suitable conformation or produces a detrimental steric hindrance for binding to the protein and elicit a potent antitubulin activity. The most potent compound bears an olefinic bridge and a non-substituted phenyl as ring B. However, all of them are less potent than the parent CA-4. It is proposed that the removal of the second methoxy group on ring A could contribute to a higher potency.

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## References and notes

- Microtubules; Hyams, J., Lloyd, C. W., Eds.; Wiley-Liss: New York, 1994.
- (a) Kanthou, C.; Tozer, G. M. *Blood* 2002, 99, 2060; (b) Vincent, L.; Kermani, P.; Young, L. M.; Cheng, J.; Zhang, F.; Shido, K.; Lam, G.; Bompais-Vincent, H.; Zhu, Z.; Hicklin, D. J.; Bohlen, P.; Chaplin, D. J.; May, C.; Rafi, S. *J. Clin. Invest.* 2005, 115, 2992.
- 3. Chaplin, D. J.; Pettit, G. R. Anticancer Res. 1999, 19, 189.
- 4. Chaplin, D. J.; Horsman, M. R.; Siemann, D. W. Curr. Opin. Invest. Drugs 2006, 7, 522.
- Cushman, M.; Nagarathnam, D.; Gopal, D.; Chakraborti,
   A. K.; Lin, C. M.; Hamel, E. J. Med. Chem. 1991, 34, 2579.
- 6. Pettit, G. R.; Singh, S. B. Can. J. Chem. 1987, 65, 2390.
- (a) Pettit, G. R.; Toki, B. E.; Herald, D. L.; Boyd, M. R.; Hamel, E.; Pettit, R. K.; Chapuis, J. C. *J. Med. Chem.* 1999, 42, 1459; (b) Shirai, R.; Takayama, H.; Nishikawa, A.; Koiso, Y.; Hashimoto, Y. *Bioorg. Med. Chem. Lett.* 1998, 8, 1997.
- 8. (a) Medarde, M.; Maya, A. B. S.; Pérez-Melero, C. *J. Enzyme Inhib. Med. Chem.* **2004**, *19*, 521, and references cited therein; (b) Tron, G. C.; Pirali, T.; Sorba, G.; Pagliai, F.; Busacca, S.; Genazzani, A. *J. Med. Chem.* **2006**, *49*, 3033, and references cited therein.
- Ravelli, R. B. G.; Gigant, B.; Curmi, P. A.; Jourdain, I.; Lachkar, S.; Sobel, A.; Knossow, M. *Nature* 2004, 428, 198.
- 10. Nam, N.-H. Curr. Med. Chem. 2003, 10, 1697.
- Kim, Y.; Arai, M. A.; Arai, T.; Lamenzo, J. O.; Dean, E. F.; Patterson, N.; Clemons, P. A.; Schreiber, S. L. J. Am. Chem. Soc. 2004, 126, 14740.
- Maya, A. B. S.; Pérez-Melero, C.; Mateo, C.; Alonso, D.;
   Fernández, J. L.; Gajate, C.; Mollinedo, F.; Peláez, R.;
   Caballero, E.; Medarde, M. J. Med. Chem. 2005, 48, 556.

- 13. Mateo, C.; López, V.; Medarde, M.; Peláez, R. *Chem. Eur. J.*, in press. doi:10.1002/chem.200700323.
- Mateo, C.; Pérez-Melero, C.; Peláez, R.; Medarde, M. J. Org. Chem. 2005, 70, 6544.
- Pettit, G. R.; Rhodes, M. R.; Herald, D. L.; Hamel, E.; Schmidt, J. M.; Pettit, R. K. J. Med. Chem. 2005, 48, 4087.
- 16. The conformations were calculated as described in reference 13. Docking attempts resulted in poses with the trimeth-oxyphenyl ring displaced from its site in tubulin, probably due to the rigid structure of the protein, which results in steric hindrance with the linker. We therefore decided to superimpose the aromatic rings of both molecules, by using the superimpose utility of Macromodel<sup>®</sup>.